

# Effect of Insulin on the Rate of Hydrogen Peroxide Generation in Mitochondria

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We studied the effect of insulin on  $H_2O_2$  generation by mitochondria in rat liver and heart. Insulin markedly increased the rate of  $H_2O_2$  generation, which was realized via short-term activation of mitochondrial succinate dehydrogenase. In terms of the Michaelis-Menten equation describing the dependence of  $H_2O_2$  generation by mitochondria on succinate concentration (succinate dehydrogenase substrate), insulin decreased the Michaelis-Menten constant and increased the maximum rate of  $H_2O_2$  generation compared to the control.

**Key Words:** *succinate dehydrogenase; mitochondrion; insulin; hydrogen peroxide; succinic acid*

Insulin receptors are regulated via reversible phosphorylation. Receptor tyrosine kinase activates, while intracellular protein tyrosine phosphatases (PTP) dephosphorylate and inactivate insulin receptors [2]. Insulin inactivates PTP and initiates cascade signal transduction in cells by stimulating the release of  $H_2O_2$  that acts as a physiological inhibitor of PTP [4,7]. NADPH oxidase acts as an insulin-sensitive source of  $H_2O_2$  in adipocytes [5]. Our previous experiments showed that the mitochondrial respiratory chain is an insulin-sensitive source of  $H_2O_2$ . Succinate dehydrogenase (SDH) plays a key role in insulin-induced generation of  $H_2O_2$  and PTP inactivation in the liver [1].

Here we studied the effect of insulin on generation of  $H_2O_2$ , a physiological inhibitor of PTP, by heart and liver mitochondria.

## MATERIALS AND METHODS

$H_2O_2$  concentration was determined by phenol red oxidation in the presence of horseradish peroxidase [6].

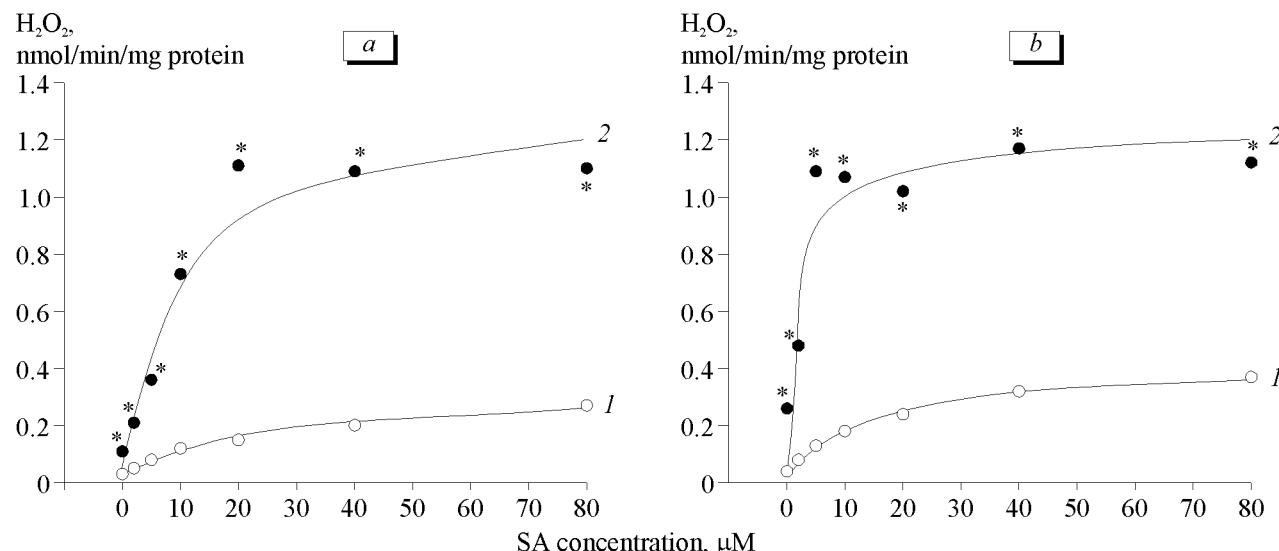
Experiments with insulin were performed on slices of the liver and heart from male Wistar rats. Slices (1 mm) were thoroughly washed with 0.15 M KCl. For evaluation of the effect of insulin on the rate of  $H_2O_2$  generation by mitochondria the slices were incubated in a medium containing 30 mM Tris-HCl (pH 7.4) in the presence (experiment) or absence of 1  $\mu$ U/ml insulin (control) at 37°C for 1 min. Mitochondria were isolated from slices by differential centrifugation in 0.25 M sucrose with 0.1 M EDTA. Nuclei were removed at 600g. Mitochondria were isolated at 10,000g. Isolated mitochondria were incubated in a medium containing 35 mM Tris-HCl (pH 7.6) and succinic acid (SA) in increasing concentrations at 37°C for 15 min. The concentration of  $H_2O_2$  in the incubation medium was measured.

The significance of differences was evaluated by Student's *t* test. The maximum rate of  $H_2O_2$  generation ( $V_m$ ) and Michaelis constant ( $K_m$ ) were calculated using regression analysis.

## RESULTS

The dependence of the initial rate of  $H_2O_2$  generation by isolated mitochondria on SA concentration (SDH substrate) was described by the Michaelis-Menten equation:

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**Fig. 1.** Rate of  $\text{H}_2\text{O}_2$  generation by heart (a) and liver (b) mitochondria as a function of succinic acid (SA) concentration: control (1) and pretreatment with insulin (2). Symbols: experimental data. Lines: estimated curves. \* $p<0.01$  compared to the control ( $n=3$ ).

$$v = V_m S / (K_m + S),$$

where  $v$  is the initial rate of  $\text{H}_2\text{O}_2$  generation, and  $S$  is SA concentration.

The rate of  $\text{H}_2\text{O}_2$  generation in mitochondria isolated from insulin-pretreated tissues was higher than in the control (substrate SA, Fig. 1). Pretreatment with insulin modified parameters of the Michaelis—Menten equation describing the kinetics of  $\text{H}_2\text{O}_2$  generation by mitochondria. Insulin increased  $V_m$  and decreased  $K_m$  compared to the control (Table 1).

These equations characterize the effect of insulin on the rate of  $\text{H}_2\text{O}_2$  generation by mitochondria in the presence of SA in physiological concentrations (1-9  $\mu\text{mol/liter}$ ) [3]. In the presence of SA in these concentrations incubation of tissues with 1  $\mu\text{U/ml}$  insulin for 1 min increased the rates of  $\text{H}_2\text{O}_2$  generation by heart and liver mitochondria by 6-8 and 6-13 times, respectively.

It should be emphasized that incubation of tissues with insulin produced only a short-term increase in the rate of  $\text{H}_2\text{O}_2$  generation by mitochondria. Increasing the time of treatment with insulin to 5 min was followed by a decrease in the rate of  $\text{H}_2\text{O}_2$  generation to the control level.

Our results indicate that insulin increases the rate of  $\text{H}_2\text{O}_2$  generation due to short-term activation of mitochondrial SDH. The molecular mechanism underlying insulin-induced activation of SDH remains unclear.

We showed for the first time that insulin increased the rate of  $\text{H}_2\text{O}_2$  generation (physiological inhibi-

**TABLE 1.** Parameters of Michaelis—Menten Equation Describing the Kinetics of  $\text{H}_2\text{O}_2$  Generation by Mitochondria

Tissue pretreatment	$V_m$ , nmol/mg protein/min	$K_m$ , $\mu\text{mol/liter}$
Heart mitochondria		
Control	0.31	17.2
Insulin, 1 $\mu\text{U/ml}$	1.32	8.6
Liver mitochondria		
Control	0.42	12.4
Insulin, 1 $\mu\text{U/ml}$	1.20	1.8

tor of PTP) due to short-term activation of mitochondrial SDH.

## REFERENCES

1. I. A. Pomytkin and O. E. Kolesova, *Byull. Eksp. Biol. Med.*, **133**, No. 6, 656-658 (2002).
2. B. J. Goldstein, F. Ahmad, W. Ding, *et al.*, *Mol. Cell Biochem.*, **182**, Nos. 1-2, 91-99 (1998).
3. G. Komaromy-Hiller, P. J. Sundquist, L. J. Jacobsen, and K. L. Nuttall, *Ann. Clin. Lab. Sci.*, **27**, No. 2, 163-168 (1997).
4. K. Mahadev, A. Zilberman, L. Zhu, and B. J. Goldstein, *J. Biol. Chem.*, **276**, No. **24**, 21,938-21,942 (2001).
5. K. Mahadev, X. Wu, A. Zilberman, *et al.*, *J. Biol. Chem.*, **276**, No. 52, 48,662-48,669 (2001).
6. E. Pick and Y. Keisari, *J. Immunol. Methods*, **38**, 161-170 (1980).
7. J. Tao, C. C. Malbon, and H. Y. Wang, *J. Biol. Chem.*, **276**, No. 31, 29,520-29,525 (2001).